

SNS COLLEGE OF PHARMACY AND HEALTH SCIENCES

Affiliated To The Tamil Nadu Dr. MGR Medical University, Chennai

Approved by Pharmacy Council of India, New Delhi.

Coimbatore -641035

COURSE NAME : PHARMACEUTICAL MICROBIOLOGY - BP303 T

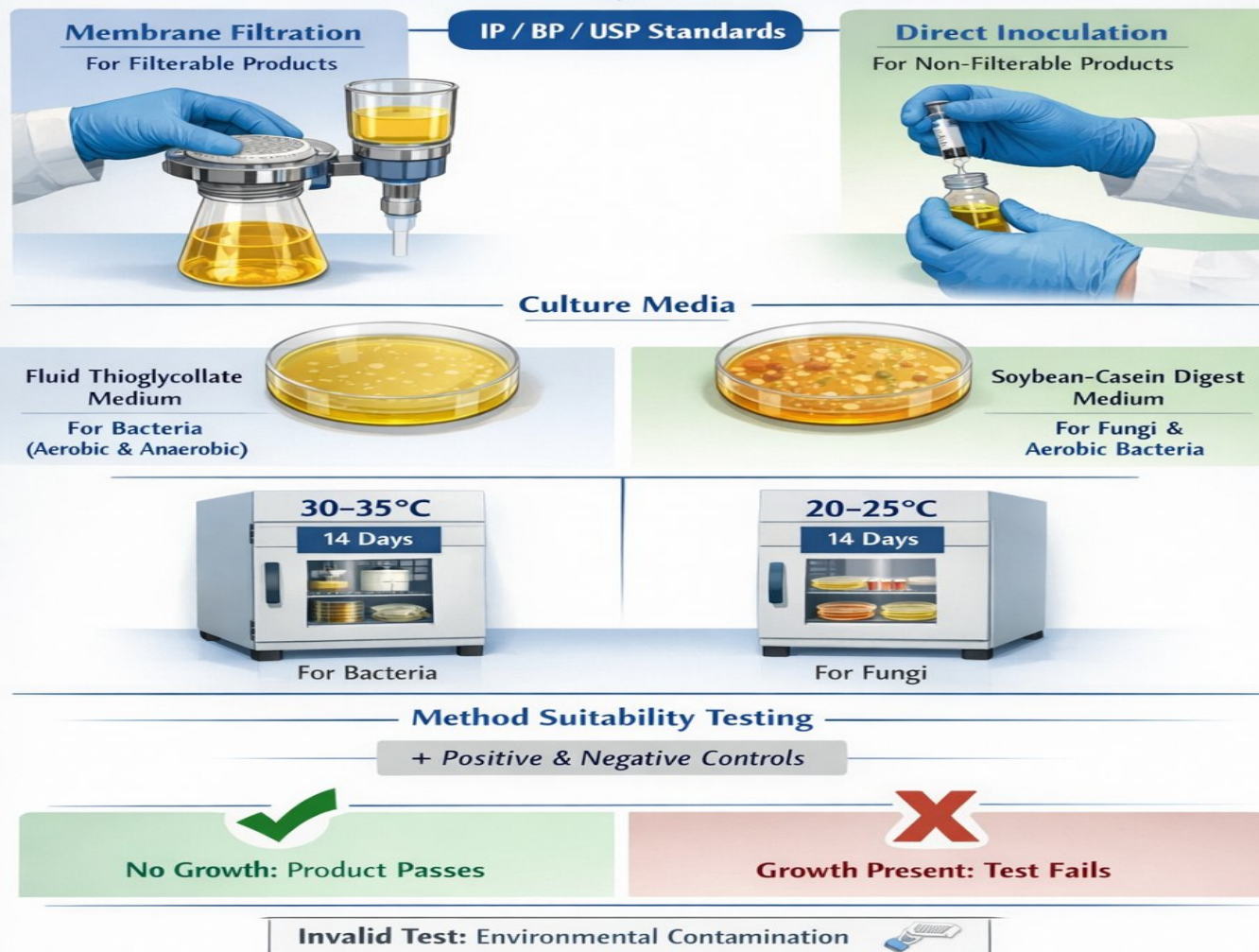
B.PHARM II YEAR / III SEM

UNIT 3

SUB TOPIC :STERILITY TESTING OF PHARMACEUTICALS

Sterility Testing Overview

Sterility testing is performed to detect viable contaminating microorganisms in sterile pharmaceutical products.



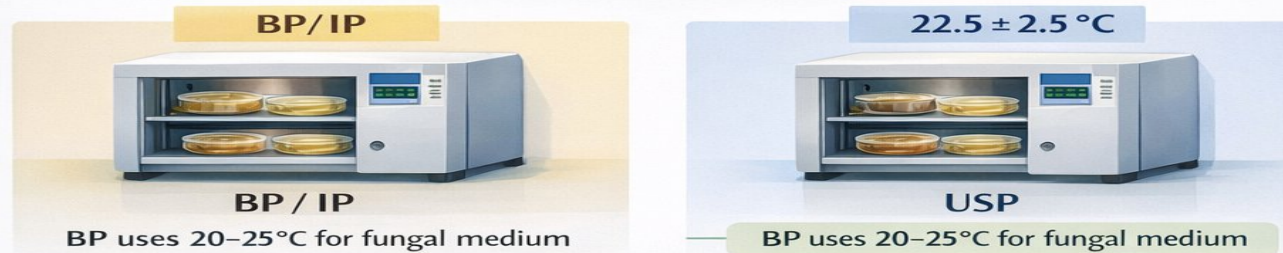
Key Differences in Sterility Testing

IP, BP, and USP are largely harmonized, but minor differences exist in alignment, fungal incubation temperatures, diluting fluids, washing membranes, and neutralizing agents used in sterility testing.

Harmonized Alignment with EP



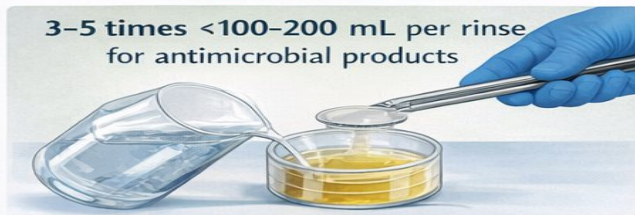
Fungal Incubation Temperature



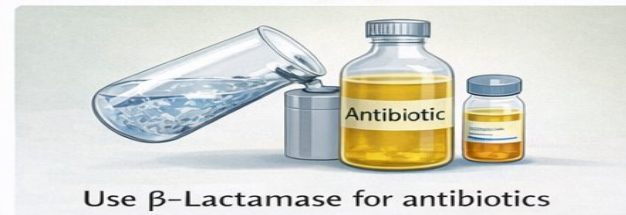
Specific Diluting Fluids (IP)



Wash Membranes



Neutralizing Agents



Membrane Filtration



Direct Inoculation

Transfer Sample Directly into Culture Media

Sample $\leq 10\%$ of Medium
Volume



Neutralize Antimicrobials

Dilution, Inactivating Agents,
or Concentrated Media



Add Emulsifiers for Greasy Products

e.g., Polysorbate 80



Shake During Incubation



Incubate for ≥ 14 Days

Observe for Growth



Product-Specific Procedures and Quantities

Quantities are minimums per medium and are similar across IP, BP, and USP (based on container content or batch size). **Use whole contents where possible;** pool from multiple containers if needed.



Batch Sample Sizes

2–10% of Batch or Minimum 2–20 Items

Depending on Batch Size & Product Type

Example Sampling Quantities:



10 Petri Dishes



2 Broth Tubes



1 Fluid Thioglycollate Bottle











For Antimicrobials:

Add Inactivators like β -Lactamase.



Minimum Quantities per Medium

Harmonized Across **IP, BP, USP**

Product Type	Minimum Quantity per Medium
 Liquids < 1 mL	Whole contents
 Liquids 1–40 mL	Half contents (≥ 1 mL)
 Liquids >40 mL but < 100 mL	20 mL
 Liquids > 100 mL	10% of contents (≥ 20 mL)
 Antibiotic liquids	1 mL
 Solids < 50 mg	Whole contents
 Solids 50–300 mg	Half contents (≥ 50 mg)
 Solids 300 mg–5 g	150 mg
 Solids > 5 g	500 mg
 Ointments/Creams (non-emulsifiable)	≥ 200 mg

Liquids (Including Ophthalmic Solutions)

Preferred: Membrane filtration

Dilute with sterile diluent (e.g., Fluid A) to ~100 mL if needed; filter directly.



For Immiscible Liquids or Suspensions

Add emulsifiers or enzymes (e.g., lecithin, penicillinase).



Ophthalmic: Treat as liquids

Use whole container or pooled amounts for non-injectables.



For Aerosols (USP/IP)

Freeze & Transfer Contents



For Vacuum Packs (IP)

Admit Sterile Air



SOLIDS (Soluble and Insoluble)

SOLUBLE

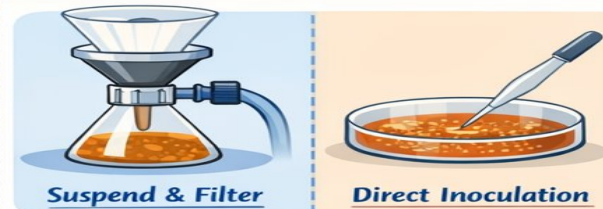
Preferred: Membrane Filtration
after dissolving in sterile solvent
(e.g. Water for Injection or Fluid A).



Filter through Membrane

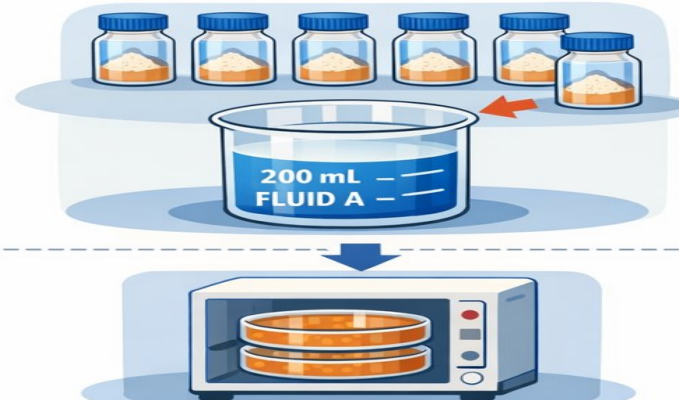
INSOLUBLE

Suspend and Filter
or Use Direct Inoculation



ANTIBIOTICS/BULKS (USP/IP)

Composite samples (e.g. 6 g from multiple containers in 200 mL Fluid A).



Incubate as Standard.

Ophthalmic and Other Non-Injectable Preparations

Treat as liquids or ointments/creams;
membrane filtration or direct inoculation.

Dilute to ~100 mL



For Ophthalmic Ointments:
Dilute 1:10 with emulsifier



Batch: 5% or minimum 2 items for ≤ 200 containers (BP/USP)



Other Sterile Products (e.g., Oils, Ointments, Creams, Devices, Sutures)

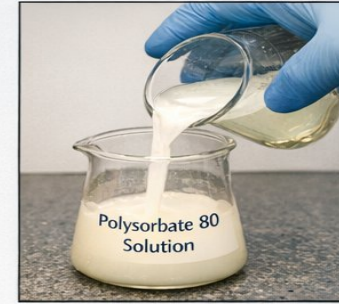
Oils / Oily Solutions



Filter Low-Viscosity



**Dilute High-Viscosity
with Isopropyl Myristate**



**Wash with
Emulsifier Solution**

Fluid B or Polysorbate 80 (10 g/L)

Use Gradual Pressure

Ointments / Creams



**Dilute to 1% in
Isopropyl Myristate
($\leq 40^{\circ}\text{C}$)**



Filter Rapidly



**Or Emulsify 1:10
for Inoculation**

Devices
(Syringes, Catheters)



Flush with ≥ 100 mL diluent
(e.g, Fluid D/B with polysorbate);
filter flushate.
For lumens, fill and immerse.

Sutures / Catgut



Direct inoculation of 3×30 cm strands
(beginning, middle, end)
in 20–150 mL medium.

Dressings / Gauze



Immerse 100–500 mg
from innermost part
or whole item.

If insoluble in isopropyl myristate, use direct inoculation (IP).

1 The Overconfident Injection



2 The Filter That Got Choked



What should you do FIRST?

- A. Increase pressure until filter cries**
- B. Change to 0.45 μm membrane**
- C. Pre-filter or dilute with suitable sterile fluid**
- D. Skip sterility testing**

Sounds like a really bad idea to me!



3 Media Identity Crisis

Two culture media are sitting
In the incubator.
One likes wai~~tn~~ weather.
the other prefers cool.

30-35°C 20-25°C

Who is who?

- A. Both like same temperature**
- B. FTM = 30-35°C, SCDM = 20-25°C**
- C. SCDM loves heat waves**
- D. Media don't care about temperature**

4 The Sneaky Microbe Party

After 7 days, the sterility test shows turbidity. The product argues:

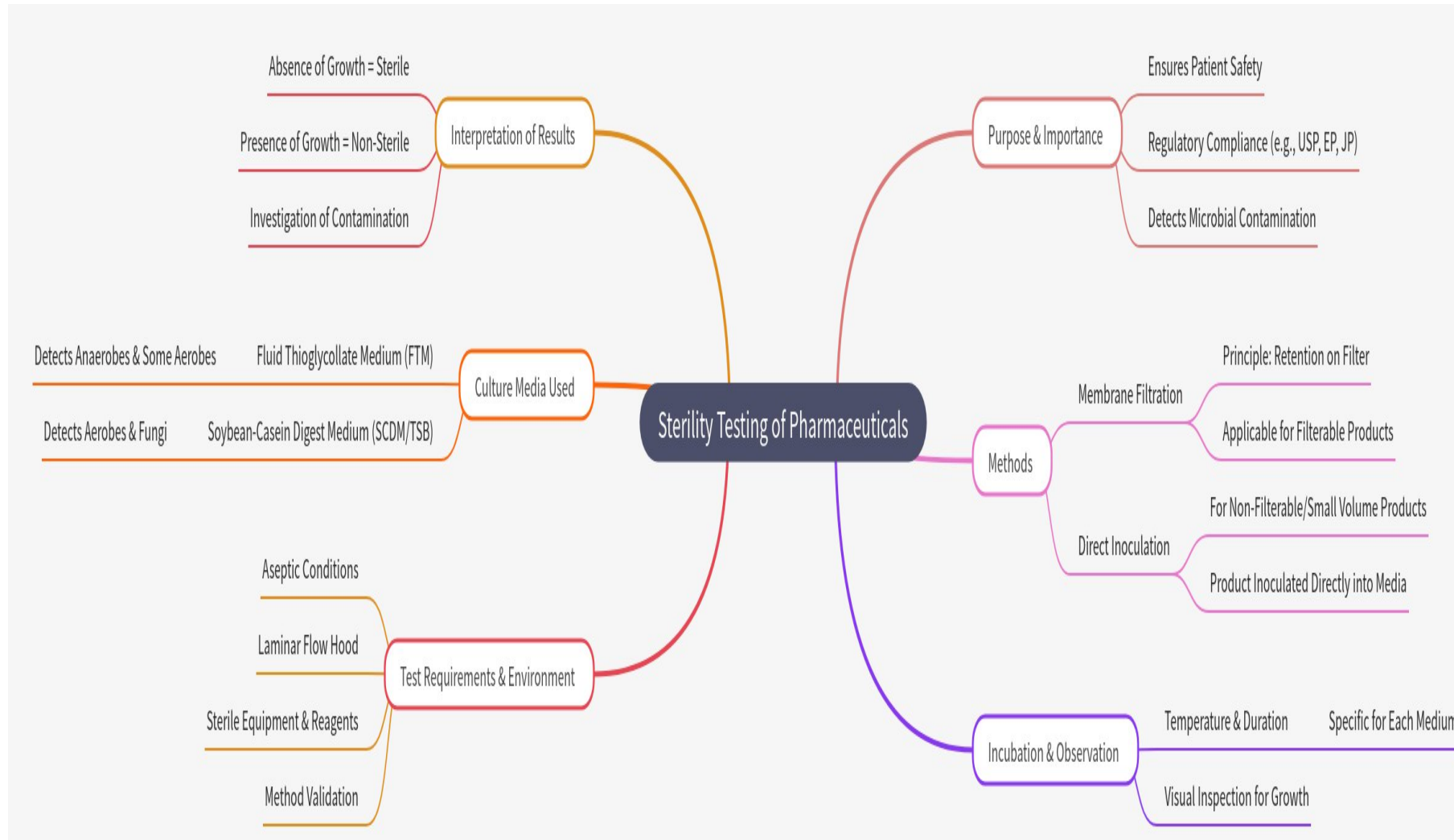
“Maybe the microbes are just visiting, not living.”

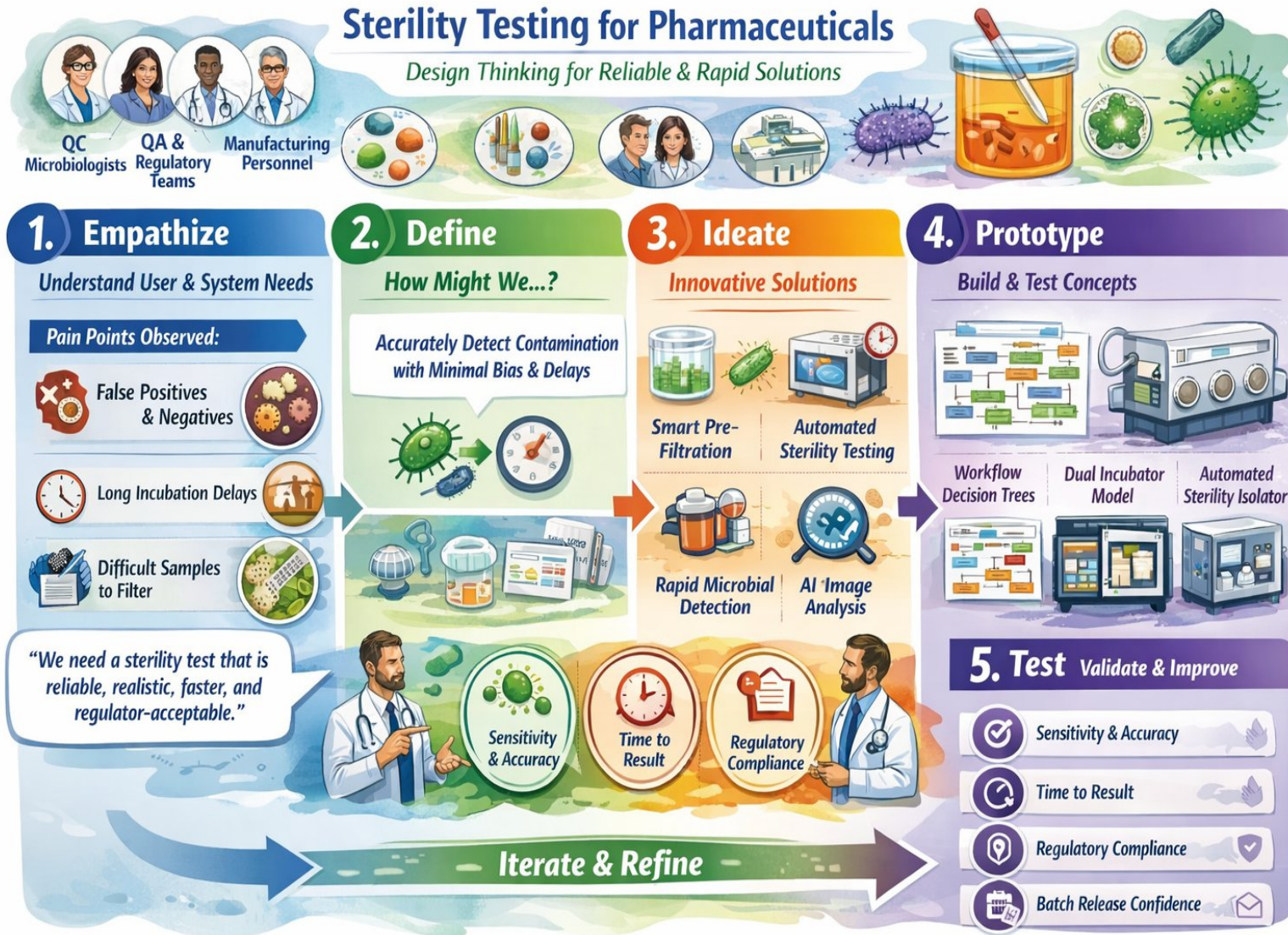


What is the correct scientific verdict?

- A. Visitors are allowed**
- B. Wait and see till 14 days**
- C. FAIL – any growth = contamination**
- D. Give microbes warning letter**







REFERENCES :

- 1.W.B. Hugo and A.D. Russel: Pharmaceutical Microbiology, Blackwell Scientific publications, Oxford London.
2. Prescott and Dunn., Industrial Microbiology, 4th edition, CBS Publishers & Distributors, Delhi.
3. Ananthanarayan : Text Book of Microbiology, Orient-Longman, Chennai

